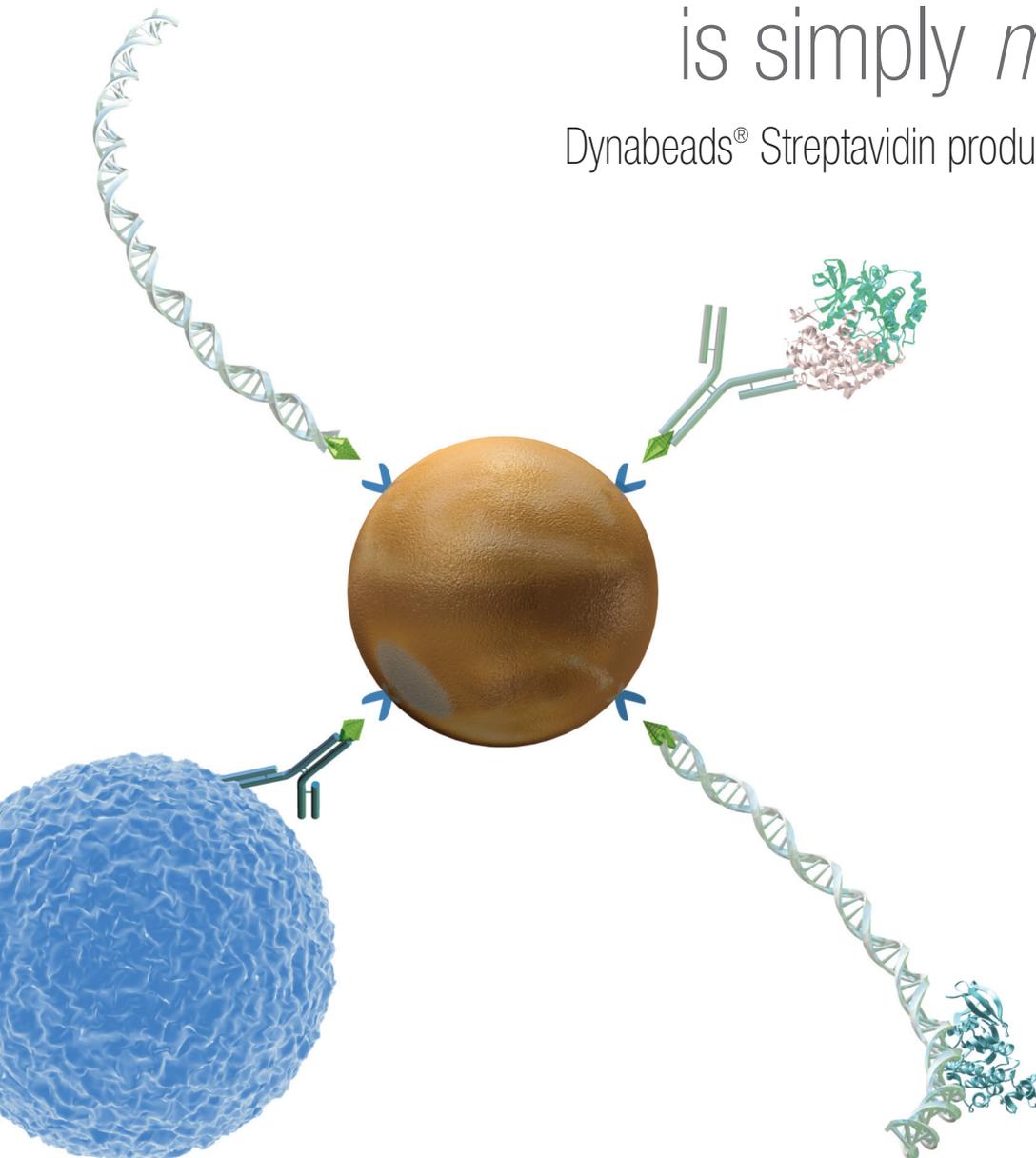




The attraction is simply *magnetisk*

Dynabeads® Streptavidin products and applications



DYNAL®



Instant capture of any biotinylated molecule

The gold standard for magnetic separations

- No centrifugation, precipitation, or columns
- In-solution reaction with rapid kinetics
- Excellent mechanical and chemical stability
- Remove variability and increase consistency

Dynabeads® Streptavidin is the gold standard for capturing, isolating, and handling biotinylated molecules. Invented in Norway and used in laboratories worldwide for more than twenty years, these groundbreaking magnetic beads are irresistibly attractive for a wide variety of applications.

The *magnetisk* quest of Dynabeads®

Magnetisk is the Norwegian word for magnetic, the property that makes Dynabeads® so attractive for a wide range of research interests, including proteomics, nucleic acid isolation, cell separation and expansion, and IVD assay development. This rapid and flexible technology makes even complicated protocols simple.

Pioneered in the 1980s by Dynal Biotech, now part of Life Technologies, Dynabeads® are based on technology developed by the late John Ugelstad, a professor of chemistry at the University of Trondheim, Norway. Ugelstad succeeded in making spherical polystyrene beads of exactly uniform size, a feat previously achieved only by NASA in the weightless conditions of space. When the uniform beads were made magnetizable, this revolutionized separation methodologies and enabled researchers to get results once

considered unattainable. Today, Dynabeads® have become the first choice among researchers for magnetic separation technology, and are used in academic and industry laboratories worldwide. They're employed on more than 25,000 IVD instruments. Some 10,000 scientific articles have been published that involve their use.

The monosized Dynabeads® provide a consistent and defined surface for the adsorption or coupling of various bioreactive molecules. Their superparamagnetism means they exhibit magnetic properties only in a magnetic field, with no residual magnetism once the field is removed. When added to a sample, Dynabeads® bind to the desired target—cells, nucleic acids, proteins, or other biomolecules. When placed in a magnetic field, bound material is rapidly and efficiently separated from the rest of the sample.

Try Dynabeads® for your next research application challenge. To learn more, please visit us at www.invitrogen.com/dynabeads.

Easy handling

Magnetic separation is surprisingly easy. No tedious centrifugation, precipitation, filtration, or columns. Magnetic handling enables easy washing, separation, and concentration of your target. Excellent dispersion abilities and the lack of magnetic remanence make Dynabeads® ideal for manual as well as automated protocols, including microfluidic systems. Depending on your specific application and target molecule, a direct or indirect capture method is applied (Figure 1).

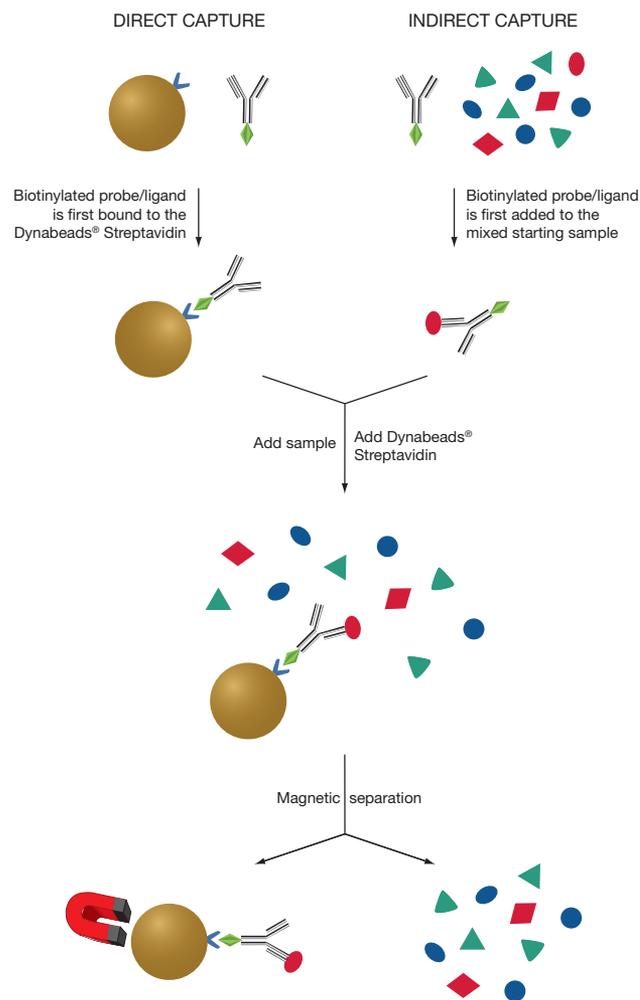


Figure 1. Direct and indirect approach for magnetic separation. In direct capture, the target-specific ligand is bound to the Dynabeads® and then added to the sample. For some applications, this enables reuse of the beads, thereby reducing costs. In indirect capture, the ligand is first allowed to bind to the target, prior to addition of Dynabeads®. This can be beneficial when the concentration of the target is low, the specific affinity is weak, or the binding kinetics are slow.



Fast and flexible

Dynabeads® Streptavidin enables instant and efficient capture of biotinylated molecules via rapid liquid-phase kinetics. This increases speed and sensitivity compared to filters and plate-based approaches (Figure 2). Dynabeads® are truly spherical and have a large surface area per volume. This ensures a high and constant binding capacity. Figure 3 highlights some selected application examples.

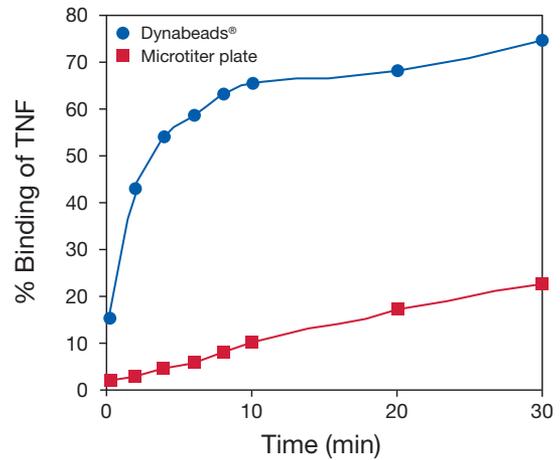


Figure 2. Dynabeads® binding kinetics are superior to traditional microtiter plates. The graph shows % binding of tumor necrosis factor (TNF) to immobilized antibody as a function of time. Courtesy of Dr. N-B Liabakk, University of Trondheim, Norway.

Sequence-specific capture

- Low-abundance cDNA from libraries
- Mutated sequences⁵
- RNA/DNA infectious agents¹
- Microsatellite enrichment¹²

Immobilized DNA/cDNA

- DNA-/RNA-binding protein isolation¹⁹
- Solid-phase DNase footprinting⁷
- Solid-phase S1 nuclease mapping⁶
- Subtractive hybridization⁸
- Differential display¹³
- Limes⁸
- 5'RACE¹⁴
- SAGE¹⁵
- TOGA¹⁶
- RAGE¹⁷

Single-stranded template

- Solid-phase sequencing
- Pyrosequencing¹⁰
- MALDI-MS³
- Probe generation⁹
- Allele-specific extension⁵
- *In vitro* mutagenesis

Specific cell isolation

- Cell culture²⁹
- Flow cytometry
- Cell-cell interactions
- Chemokine and immunological assays²⁷
- Bacterial pathogen detection³⁰
- Molecular analyses²⁸

Protein purification

- Intact protein complexes
- Active enzymes³⁶
- Immunoprecipitation²²
- Protein interaction studies
- Protein depletion
- SDS-PAGE
- MALDI-TOF¹⁹

Immunoassays

- Competitive³⁴/noncompetitive²³
- Homogeneous³³/heterogeneous²⁴
- Sandwich assays²

Biopanning

- Phage display^{34,36}
- Cell-based screening³⁵
- SELEX³²
- Affibody® selection³¹
- Drug screening²³

Figure 3. Selected applications using Dynabeads® Streptavidin.

Robust and gentle

Dynabeads® Streptavidin features excellent mechanical and chemical stability. No iron leakage. No inhibition of enzyme activity. The monolayer of recombinant streptavidin ensures that the vast majority of biotin-binding sites are left sterically available for binding.

The technology is also extremely gentle, allowing isolation of proteins as well as large or unstable complexes. The native state of proteins is preserved, and fragile cells remain viable. Flexible volumes may be used, enabling isolation of low-abundance molecules. A high signal-to-noise ratio also contributes to increased sensitivity.

Absolute reproducibility

All Dynabeads® are produced with full control of parameters such as bead size, surface area, iron content, and magnetic mobility. The absence of excess physically-adsorbed streptavidin ensures negligible leakage, and secures a minimal batch-to-batch variation. The uniform characteristics and unique reproducibility within (CV <3%) and between batches reduce costs associated with quality control testing (Figure 4). Whether for your research project or IVD testing activities, you can rely on the consistent performance of Dynabeads®.

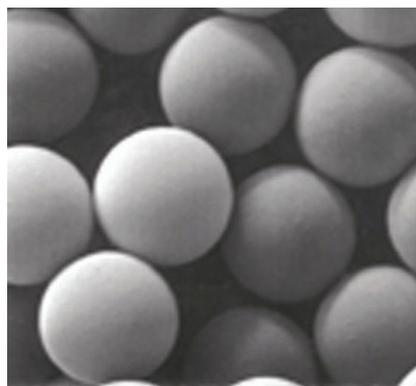


Figure 4. Monosized superparamagnetic Dynabeads®. Each bead has an even dispersion of magnetic material, encased within a thin polymer shell. This provides a specific and defined surface for binding your ligand. The true uniformity of all beads within each batch (typical CV <3%) provides consistent physical and chemical properties. Unique batch-to-batch reproducibility (typical CV <5%) secures reproducibility and quality of results.



Choose your favorite

Four different types of Dynabeads® Streptavidin are available (Table 1). Your choice should be guided by your sample and target properties, buffers and solutions applied, and specific downstream application needs.

Dynabeads® M-280 Streptavidin and Dynabeads® MyOne™ Streptavidin T1 are commonly used for protein and nucleic acid appli-

cations. Dynabeads® M-270 Streptavidin and Dynabeads® MyOne™ Streptavidin C1 are preferred for nucleic acid protocols involving high chaotropic salt concentrations, for immunoassays involving small biotinylated antigens, and applications incompatible with BSA. The smaller MyOne™ beads offer increased binding capacity and slower sedimentation rate, making them ideal for automated applications.

Table 1. An overview of the different Dynabeads® Streptavidin, and their qualities based on selected applications.

| Product | Binding capacities | Characteristics and properties | Ideal for |
|-----------------------------------|---|---|---|
| Dynabeads® M-280 Streptavidin | Free biotin: 650–900 pmol/mg beads Biotinylated Ig: Up to 10 µg/mg beads | <ul style="list-style-type: none"> Hydrophobic bead surface Based on tosylactivated beads Diameter: 2.8 µm Size distribution: CV <3% BSA as blocking protein Isoelectric point: pH 5.0 Low charge (–10 mV (at pH 7)) Iron content (ferrites): 12% (17%) | <ul style="list-style-type: none"> Immunoassays Purification of DNA-/RNA-binding proteins Protein purification Phage display Biopanning Cell isolation |
| Dynabeads® MyOne™ Streptavidin T1 | Free biotin: 1,100–1,700 pmol/mg beads Biotinylated Ig: Up to 20 µg/mg beads | <ul style="list-style-type: none"> Hydrophobic bead surface Based on tosylactivated beads Diameter: 1.05 µm Size distribution: CV <3% BSA as blocking protein Isoelectric point: pH 5.0 Low charge (–10 mV (at pH 7)) Iron content (ferrites): 26% (37%) Low sedimentation rate and faster reaction kinetics compared to M-280/M-270 beads | <ul style="list-style-type: none"> Immunoassays Purification of DNA-/RNA-binding proteins Protein purification Phage display Biopanning Cell isolation Well suited for automated applications |
| Dynabeads® M-270 Streptavidin | Free biotin: ≥950 pmol/mg beads Biotinylated Ig: Up to 10 µg/mg beads | <ul style="list-style-type: none"> Hydrophilic bead surface Based on carboxylic acid beads Diameter: 2.8 µm Size distribution: CV <3% No blocking proteins used Isoelectric point: pH 4.5 High charge (–50 mV (at pH 7)) Iron content (ferrites): 14% (20%) Low aggregation of beads in high-salt solutions | <ul style="list-style-type: none"> Sequence-specific DNA/RNA capture in nucleic acid research Protocols that require GTC lysis or high salt concentrations Preparation of single-stranded DNA Immunoassays with hydrophobic targets |
| Dynabeads® MyOne™ Streptavidin C1 | Free biotin: ≥2,500 pmol/mg beads Biotinylated Ig: Up to 20 µg/mg beads | <ul style="list-style-type: none"> Hydrophilic bead surface Based on carboxylic acid beads Diameter: 1.05 µm Size distribution: CV <3% No blocking proteins used Tween 20 in the buffer Isoelectric point: pH 5.2 Medium charge (–35 mV (at pH 7)) Iron content (ferrites): 26% (37%) Low sedimentation rate and faster reaction kinetics compared to M-280/M-270 beads Low aggregation | <ul style="list-style-type: none"> Sequence-specific DNA/RNA capture in nucleic acid research Preparation of single-stranded DNA High-throughput nucleic acid clean-up protocols Sample preparation of proteins for mass spectrometry Well suited for automated applications |

Selected references

Automated protocols:

1. Meng Q et al. (2001) Automated multiplex assay system for simultaneous detection of hepatitis B virus DNA, hepatitis C virus RNA and human immunodeficiency virus type 1 RNA. *J Clin Microbiol* 39(8):2937–2945.
2. Pollock GS et al. (2001) Effects of early visual experience and diurnal rhythms on BDNF mRNA and protein levels in the visual system, hippocampus and cerebellum. *J Neurosci* 21(11):3923–3931.

Single-stranded DNA templates:

3. von Wintzingerode F et al. (2002) Base-specific fragmentation of amplified 16S rRNA genes analyzed by mass-spectrometry: A tool for rapid bacterial identification. *Proc Natl Acad Sci U S A* 99(10):7039–7044.
4. Pourmand N et al. (2002) Multiplex pyrosequencing. *Nucleic Acids Res* 30(7):e31.
5. Lindblad-Toh K et al. (2000) Large-scale discovery and genotyping of single nucleotide polymorphisms in the mouse. *Nat Genet* 24:381–386.
6. Dziembowski A et al. (2001) Analysis of 3' and 5' ends of RNA by solid-phase S1 nuclease mapping. *Anal Biochem* 294:87–89.
7. Fletcher TM et al. (2002) Structure and dynamic properties of a glucocorticoid receptor-induced chromatin transition. *Mol Cell Biol* 20(17):6466–6475.
8. Hansen-Hagge TE et al. (2001) Identification of sample-specific sequences in mammalian cDNA and genomic DNA by the novel ligation-mediated subtraction (Limes). *Nucleic Acids Res* 29(4):e20.
9. Beulieu M et al. (2001) PCR candidate region mismatch scanning: adaptation to quantitative, high-throughput genotyping. *Nucleic Acids Res* 29(5):1114–1124.
10. Leunissen ME et al. (2009) Switchable self-protected attractions in DNA-functionalized colloids. *Nature Mater* 8:590–595.
11. Helena Persson H et al. (2009) The non-coding RNA of the multidrug resistance-linked vault particle encodes multiple regulatory small RNAs. *Nature Cell Biol* 11(10):1268–1271.

Sequence-specific capture:

12. Refseth UH et al. (1997) Hybridization capture of microsatellites directly from genomic DNA. *Electrophoresis* 18(9):1519–1523.
13. Kornmann B et al. (2001) Analysis of circadian liver gene expression by ADDER, a highly sensitive method for the display of differentially expressed mRNAs. *Nucleic Acids Res* 29(11):e51.
14. Schramm G et al. (2000) A simple and reliable 5'-RACE approach. *Nucleic Acids Res* 28(22):e96.
15. Velculescu VE et al. (1995) Serial analysis of gene expression. *Science* 270(5235):484–487.
16. Sutcliffe JG et al. (2000) TOGA: An automated parsing technology for analyzing expression of nearly all genes. *Proc Natl Acad Sci U S A* 97(5):1976–1981.
17. Wang A et al. (1999) Rapid analysis of gene expression (RAGE) facilitates universal expression profiling. *Nucleic Acids Res* 27(23):4609–4618.
18. Gnirke A et al. (2009) Solution hybrid selection with ultra-long oligonucleotides for massively parallel targeted sequencing. *Nature Biotechnol* 27(2):182–189.

Nucleic acid-binding proteins:

19. Nordhoff E et al. (1999) Rapid identification of DNA-binding proteins by mass spectrometry. *Nat Biotechnol* 17:884–888.
20. Noguchi E et al. (2009) A Crohn's disease-associated NOD2 mutation suppresses transcription of human IL10 by inhibiting activity of the nuclear ribonucleoprotein hnRNP-A1. *Nature Immunol* 10(5):471–479.
21. Li D et al. (2009) Down-Regulation of MHC Class II Expression through Inhibition of CIITA Transcription by Lytic Transactivator Zta during Epstein-Barr Virus Reactivation. *J Immunol* 182(4):1799–1809.

Protein purification:

22. O'Reilly FM et al. (2002) FKBP12 modulation of the binding of the skeletal ryanodine receptor onto the II-III loop of the dihydropyridine receptor. *Biophys J* 82:145–155.
23. Chao S-H and Price DH (2001) Flavopiridol inactivates P-TEFb and blocks most RNA polymerase II transcription in vivo. *J Biol Chem* 276(34):31793–31799.
24. deBaar MP et al. (1999) Detection of human immunodeficiency virus type I nucleocapsid protein p7 in vitro and in vivo. *J Clin Microbiol* 37(1):63–67.
25. Lehrbach NJ et al. (2009) LIN-28 and the poly(U) polymerase PUP-2 regulate let-7 microRNA processing in *Caenorhabditis elegans*. *Nat Struct Mol Biol* 16(10):1016–1020.
26. Cook PJ et al. (2009) Tyrosine dephosphorylation of H2AX modulates apoptosis and survival decisions. *Nature* 458(7243):591–596.

Specific cell isolation:

27. Konishi Y et al. (2002) Isolation of living neurons from human elderly brains using the immunomagnetic sorting DNA-linker system. *Am J Pathol* 161(5):1567–1576.
28. Fahrer AM et al. (2001) Attributes of $\gamma\delta$ intraepithelial lymphocytes as suggested by their transcriptional profile. *Proc Natl Acad Sci U S A* 98(18):10261–10266.
29. Johansen M et al. (2002) An investigation of methods for enriching trophoblasts from maternal blood. *Prenat Diagn* 15:921–931.
30. Sun W et al. (2001) Food-borne pathogens. Use of bioluminescent *Salmonella* for assessing the efficiency of constructed phage-based biosorbent. *J Ind Microbiol Biotech* 27:126–128.

Biopanning:

31. Nord K et al. (2001) Recombinant human factor VIII-specific affinity ligands selected from phage-displayed combinatorial libraries of protein A. *Eur J Biochem* 268:4269–4277.
32. Biroccio A et al. (2002) Selection of RNA aptamers that are specific and high-affinity ligands of the hepatitis C virus RNA-dependent RNA polymerase. *J Virol* 76(8):3688–3696.
33. Legendre D et al. (1999) Engineering a regulatable enzyme for homogeneous immunoassays. *Nat Biotechnol* 17:67–72.
34. Lev A et al. (2002) Isolation and characterization of human recombinant antibodies endowed with the antigen-specific, major histocompatibility complex-restricted specificity of T cells directed toward the widely expressed tumor T cell epitopes of the telomerase catalytic subunit. *Cancer Res* 62(11):3184–3194.
35. Cumbers SJ et al. (2002) Generation and iterative affinity maturation of antibodies in vitro using hypermutating B cell lines. *Nat Biotechnol* 20(11):1129–1134.
36. Demartis S et al. (1999) A strategy for the isolation of catalytic activities from repertoires of enzymes displayed on phage. *J Mol Biol* 286:617–633.

Ordering information

| Products | Quantity | Cat. No. |
|---|---|-----------------|
| Dynabeads® M-280 Streptavidin | 2 mL | 112-05D |
| 2.8 µm magnetic beads with covalently coupled recombinant Streptavidin and a hydrophobic surface | 10 mL | 112-06D |
| | 100 mL | 602-10 |
| Dynabeads® MyOne™ Streptavidin T1 | 2 mL | 656-01 |
| 1 µm magnetic beads with covalently coupled recombinant Streptavidin and a hydrophobic surface | 10 mL | 656-02 |
| | 100 mL | 656-03 |
| Dynabeads® M-270 Streptavidin | 2 mL | 653-05 |
| 2.8 µm magnetic beads with covalently coupled recombinant Streptavidin and a hydrophilic surface | 10 mL | 653-06 |
| Dynabeads® MyOne™ Streptavidin C1 | 2 mL | 650-01 |
| 1 µm magnetic beads with covalently coupled recombinant Streptavidin and a hydrophilic surface | 10 mL | 650-02 |
| Dynabeads® Streptavidin Trial Kit | 4 x 1 mL | 658-01D |
| Contains 1 mL each of the four bead-types listed above | | |
| Dynal® kilobaseBINDER™ Kit | 1 kit | 601-01 |
| Contains 1 mL Dynabeads® M-280 Streptavidin, Binding and Washing Solution sufficient for 200 isolations | | |
| Dynabeads® Biotin Binder | 5 mL | 110-47 |
| For cell isolation or depletion, using your own biotinylated antibody | | |
| CELLlection™ Biotin Binder Kit— | 5 mL | 115-33D |
| For positive cell isolation and detachment, using your own biotinylated antibody | | |
| Related products | Quantity | Cat. No. |
| DynaMag™ magnets | See www.invitrogen.com/magnets for magnet recommendations | |
| HulaMixer™ Sample Mixer | Holds 0.5 mL–50 mL tubes | 159-20D |

Learn more and order your Dynabeads® Streptavidin today at www.invitrogen.com/dynabeads.



DYNAL® has pioneered magnetic separation technologies for biological discovery that are both simple and highly reproducible. Based on their patented superparamagnetic, monodisperse beads, Dynabeads® technologies represent a superior paradigm for cell and biomolecule separation in a wide range of basic and clinical research applications, diagnostic assays, and therapeutic protocols.



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